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### Reports

## A Mechanism of Extreme Growth and **Reliable Signaling in Sexually Selected Ornaments and Weapons**

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Many male animals wield ornaments or weapons of exaggerated proportions. We propose that increased cellular sensitivity to signaling through the insulin/insulinlike growth factor (IGF) pathway may be responsible for extreme growth of these structures. We document how rhinoceros beetle horns, a sexually selected weapon, are more sensitive to nutrition and more responsive to perturbation of the insulin/IGF pathway than other body structures. We then illustrate how enhanced sensitivity to insulin/IGF signaling in a growing ornament or weapon would cause heightened condition-sensitivity and increased variability in expression among individuals—critical properties of reliable signals of male quality. The possibility that reliable signaling arises as a byproduct of the growth mechanism may explain why trait exaggeration has evolved so many different times in the context of sexual selection.

The most elaborate male ornaments and weapons of sexual selection grow to exaggerated proportions (Fig. 1), especially in the largest and best-conditioned individuals. The size and conspicuousness of these traits make them likely candidates for intraspecific signals, used either by males to assess the size, condition, or status of rival males, or by females to assess the relative genetic quality of potential mates (1, 2). Not only are exaggerated traits easy to observe, they are unusually reliable signals of individual male quality (2-4) as their growth tends to be more sensitive to the nutritional histories and physiological conditions of individuals than is the growth of other traits (5–7). Exaggerated structures also tend to be more variable in their expression than other morphological structures (8-10). Hyper-variability in trait size can amplify otherwise subtle differences in the body size or condition of males, further enhancing the utility of these traits as signals. Combined, these structural characteristics - extreme size, heightened condition-sensitivity, and hyper-variability among individuals – are the foundation for 'handicap' and 'good genes' models of sexual selection and a central tenet of modern theories of sexual selection and animal communication (2-4, 11-15). We offer a developmental explanation for this phenomenon. We suggest the evolution of trait exaggeration involves increased sensitivity to insulin/IGF signaling within a growing structure, and we show why such a change in mechanism should also confer both heightened condition sensitivity and hyper-variablity to expression of the trait (Figure 1B).

Insulin and IGFs are essential regulators of tissue growth and body size (16). Circulating levels of insulin and IGFs are sensitive to nutrition, as well as stress and infection, and the insulin/IGF pathway has emerged as the central mechanism integrating physiological condition with growth in multicellular animal taxa. Insulin and IGF levels within a growing animal reflect the nutritional state and physiological condition of that individual, and circulating levels of these signals modulate tissue growth via the insulin receptor pathway in a graded, or dose-dependent

manner. Within an individual, growth will speed up or slow down in response to changes in nutritional or physiological state because of the action of this pathway. Across individuals, growth will differ between high-condition and low-condition individuals, resulting in population-level variation in body and trait sizes. Low-condition individuals have lower levels of these signals than higher condition individuals, and, as a result, they experience slower rates and lower overall amounts of tissue growth.

As long as the various organs and body parts (e.g., legs, eyes, wings) exhibit similar sensitivities to insulin/IGF signaling (17), their sizes will scale proportionally from individual to individual (18–21). But some traits deviate in their responsiveness to these  $\mathbb{Q}$ signals, profoundly affecting the amount and nature of their growth. Genitalia are insensitive to circulating insulin/IGF signals in Drosophila (20, 21). As a result, their growth is unre-sponsive to environmental conditions, such as nutrition, and genitalia size is largely invariant among individuals. In contrast, wings exhibit sensitivity to insulin/IGF signaling typical of the rest of the body; wing growth is sensitive. 21). As a result, their growth is unre-

to larval nutrition, and wing sizes scale isometrically with amongindividual variation in body size (21).

We predicted that increased sensitivity to the insulin/IGF pathway might be a mechanism leading to the evolution of extreme growth in showy ornaments and weapons of sexual selection. In our model, individual males differ in their physiological state as a result of differences in their status, nutritional state, competitive ability, and/or health (parasite or pathogen loads), translating into among-individual variation in exposed to insulin/IGF signals, and the sensitivity of cells within each arows would determine both the how much each trait how much each trait grew. Just as wings are more sensitive to insulin/IGF signaling than genitalia in Drosophila (20, 21), so we predicted that exaggerated ornaments or weapons of sexual selection would be even more sensitive to insulin/IGF signaling than wings or other nonsexually-selected body parts (Fig. 1B).

Male rhinoceros beetles (Trypoxylus dichotomus) wield a forked horn on their heads. During growth, horns in this species are more sensitive to larval nutrition than other body parts (wings, genitalia), and, among adult males, horn size is hyper-variable, ranging from tiny bumps to exaggerated structures two thirds the length of a male's body (22). We tested whether growing rhinoceros beetle horns were more sensitive to insulin/IGF signaling than wings or genitalia using RNA interference to perturb transcription of the insulin receptor (InR). Developing larvae were injected with a 398bp fragment of dsRNA of T. dichotomus InR as they commenced their transition from larval feeding to gut purge (the onset of the prepupal period and the beginning of metamorphosis). At this time all growth in overall body size had ceased, but adult structures (including genitalia, wings, and horns) were still growing. Thus, any effects of manipulation of insulin/IGF signaling would be visible as

reductions to genitalia, wing, or horn size relative to overall body size. If the evolution of exaggerated horn size resulted in part from an increase in cellular sensitivity to insulin/IGF signaling, then horns should be more sensitive than wings to perturbation of the activity of this pathway. We also predicted that genitalia would be relatively insensitive to pathway perturbation (sensu 20, 21).

Injections significantly reduced InR transcript abundances for 48 hours near the end of the period of trait growth (i.e., before InR transcript abundance normally drops in these tissues; Fig. 2A-C). After metamorphosis was completed, we compared morphologies of treated and control animals. Genitalia did not respond to experimental perturbation of *InR* pathway activity (Wald statistic = 0.1245, 1 degree of freedom, p = 0.724; Fig. 2D). Wings, which exhibit nutrition-sensitive growth patterns typical of the majority of metric traits (e.g., eyes, legs, elytra, etc.), showed a significant reduction in size of ~ 2% (Wald statistic = 8.976, 1 df, p = 0.003; Fig. 2E). In contrast, male horns, the structures most sensitive to nutrition, were reduced by ~ 16% relative to controls (Wald statistic = 68.37, 1 df, p < 0.0001; Fig. 2F, G). Using response to InR knockdown as a metric, male horns were eight times more sensitive to insulin/IGF signaling than wings, consistent with our model for the evolution of disproportionate or exaggerated weapon size from enhanced tissue-specific sensitivity to the insulin/IGF pathway.

A growing body of research now implicates insulin/IGF signaling in the development of extreme animal structures (23). Insulin/IGF signaling is an ancient and conserved physiological pathway that has coupled rates of cell proliferation with available nutrients for at least 500 million years, and we suggest that this pathway has been co-opted repeatedly in lineages experiencing strong sexual selection to yield disproportionate growth in signaling structures. The insulin/IGF pathway would likely have controlled the rate of growth of these structures already; increased cellular sensitivity to these signals would therefore be an easy route to the evolution of accelerated growth if the structure came under directional sexual selection for increased size.

But such a route to exaggeration would only generate exaggerated trait sizes in high-condition individuals because low-condition individuals would have low circulating levels of insulin/IGF signals and attenuated rates of tissue and body growth. The same mechanism stimulating increased trait growth in high quality individuals would also repress trait growth in low quality individuals (Fig. 1B). This means that whenever exaggerated ornament or weapon size arises due to an increase in trait-specific sensitivity to insulin/IGF signaling, then the exaggerated trait should also show enhanced (or 'heightened') condition-sensitive expression and higher relative variability in trait size between low- and high-condition individuals (as compared to other, non-exaggerated, traits). Signal reliability would be an intrinsic property of these structures because of the developmental mechanism regulating their growth.

Theoretical considerations of sexual selection and animal signaling argue that escalated evolution of signals is most likely when signals are reliable, and it is difficult or impossible for low quality males to "cheat" by producing full-sized structures (Fig. 3). Signal reliability can be evolutionarily stable under two sets of conditions: either the signal is sufficiently costly to produce or wield that it is not cost-effective for low quality individuals to cheat ('handicap' signals), or the signal is intrinsically unfakable ('index' signals, 'good genes' signals) (2-4, 11-13, 24-33). The largest ornaments and weapons are generally assumed to be handicap signals of male quality, where the cost of these structures enforces signal reliability (2-4, 24-33). However, for even the largest of structures, the process of escalation must have started when these structures were small, and at that early stage, these costs would likely have been minimal. Moreover, several recent studies of exaggerated male ornaments and weapons have failed to find significant costs (34, 35), forcing a reconsideration of the question: why don't low quality males cheat?

We suggest that exaggerated animal structures may be unfakable signals of quality because of the developmental mechanism responsible for their accelerated growth. If true, then our hypothesis of 'intrinsic reliability' could help explain why so many different signal traits embark on an evolutionary trajectory of bigger and bigger size. We suggest that whenever receivers responded to variation in insulin/IGF-sensitive structures, they fared relatively well due to the intrinsic reliability of these traits as signals of underlying male quality. As these traits became larger under selection, their utility as signals would have increased, enhancing the benefits to receivers and accelerating the rate of signal evolution still further. Once these structures become large enough to be costly, they may also act as handicap signals and costs could contribute to signal reliability (Fig. 3). However, as long as the traits exhibit heightened sensitivity to insulin/IGF signals, costs may not be necessary for signal reliability (36). This means that subsequent evolution of compensatory structures alleviating costs to the signaling males (37) need not undermine the reliability of these traits as signals and could explain why some exaggerated sexually selected structures function as reliable signals even when no discernable costs are apparent (34, 35).

### **References and Notes**

- C. Darwin, *The Descent of Man and Selection in Relation to Sex* (Random House, Modern Library, New York, 1871).
- J. W. Bradbury, S. L. Vehrencamp, Principles of Animal Communication (Sinauer, Sunderland, MA, 2011).
- 3. J. Maynard Smith, D. Harper, *Animal Signals* (Oxford Univ. Press, Oxford, 2004).
- W. A. Searcy, S. Nowicki, The Evolution of Animal Communication: Reliability and Deception in Signaling Systems (Princeton Univ. Press, Princeton, 2005).
- S. Cotton, K. Fowler, A. Pomiankowski, Condition dependence of sexual ornament size and variation in the stalk-eyed fly *Cyrtodiopsis dalmanni* (Diptera: Diopsidae). *Evolution* 58, 1038 (2004). <u>Medline</u>
- R. Bonduriansky, L. Rowe, Sexual selection, genetic architecture, and the condition dependence of body shape in the sexually dimorphic fly *Prochyliza xanthostoma* (Piophilidae). *Evolution* 59, 138 (2005). <u>Medline</u>
- R. J. Knell, N. Fruhauf, K. A. Norris, Conditional expression of a sexually selected trait in the stalk-eyed fly *Diasemopsis aethiopica*. *Ecol. Ent.* 24, 323 (1999). doi:10.1046/j.1365-2311.1999.00200.x
- R. V. Alatalo, J. Höglund, A. Lundberg, Patterns of variation in tail ornament size in birds. *Biol. J. Linn. Soc. Lond.* 34, 363 (1988). <u>doi:10.1111/j.1095-8312.1988.tb01969.x</u>
- S. Fitzpatrick, Patterns of morphometric variation in birds' tails: Length, shape and variability. *Biol. J. Linn. Soc. Lond.* 62, 145 (1997). <u>doi:10.1111/j.1095-8312.1997.tb01619.x</u>
- J. J. Cuervo, A. P. Møller, The allometric pattern of sexually size dimorphic feather ornaments and factors affecting allometry. *J. Evol. Biol.* 22, 1503 (2009). doi:10.1111/j.1420-9101.2009.01758.x Medline
- R. A. Johnstone, Sexual selection, honest advertisement and the handicap principle: Reviewing the evidence. *Biol. Rev. Camb. Philos. Soc.* 70, 1 (1995). <u>doi:10.1111/j.1469-185X.1995.tb01439.x Medline</u>
- L. Rowe, D. Houle, The lek paradox and the capture of genetic variance by condition-dependent traits. *Proc. Biol. Sci.* 263, 1415 (1996). <u>doi:10.1098/rspb.1996.0207</u>
- Y. Iwasa, A. Pomiankowski, Good parent and good genes models of handicap evolution. J. Theor. Biol. 200, 97 (1999). <u>doi:10.1006/jtbi.1999.0979</u> <u>Medline</u>
- S. Cotton, K. Fowler, A. Pomiankowski, Do sexual ornaments demonstrate heightened condition-dependent expression as predicted by the handicap hypothesis? *Proc. Biol. Sci.* 271, 771 (2004). <u>doi:10.1098/rspb.2004.2688</u> <u>Medline</u>
- R. Bonduriansky, The evolution of condition-dependent sexual dimorphism. Am. Nat. 169, 9 (2007). doi:10.1086/510214 Medline
- 16. A more complete description of this pathway and references are provided in the Supplementary Online Material (SOM).
- 17. For this paper we define tissue sensitivity as the extent to which variations in the level of hormone signal influence the rate of cell proliferation via activity of the insulin/IGF pathway. Insensitive tissues grow to roughly the same final

size regardless of circulating insulin/IGF levels, whereas the amounts of growth of sensitive tissues are strongly regulated by signal levels. Tissue sensitivity is often equated with receptor density. However, in this case, altered expression of any number of downstream genes in the pathway could change the responsiveness of a tissue to insulin/IGF signals. Indeed, in the best-studied example to date, reduced insulin-sensitivity in a specific tissue (genitalia) in *Drosophila* resulted from lowered levels of expression of a "downstream" element of the insulin-signaling pathway, *FOXO*, and not from tissue-differences in expression of the insulin receptor (21).

- A. W. Shingleton, W. A. Frankino, T. Flatt, H. F. Nijhout, D. J. Emlen, Size and shape: The developmental regulation of static allometry in insects. *Bioessays* 29, 536 (2007). doi:10.1002/bies.20584 Medline
- A. W. Shingleton, C. K. Mirth, P. W. Bates, Developmental model of static allometry in holometabolous insects. *Proc. Biol. Sci.* 275, 1875 (2008). <u>doi:10.1098/rspb.2008.0227</u> Medline
- A. W. Shingleton, J. Das, L. Vinicius, D. L. Stern, The temporal requirements for insulin signaling during development in *Drosophila*. *PLoS Biol.* 3, e289 (2005). doi:10.1371/journal.pbio.0030289 Medline
- H. Y. Tang, M. S. B. Smith-Caldas, M. V. Driscoll, S. Salhadar, A. W. Shingleton, *FOXO* regulates organ-specific phenotypic plasticity in *Drosophila*. *PLoS Genet.* 7, e1002373 (2011). doi:10.1371/journal.pgen.1002373 Medline
- 22. Results, as well as all methods for this paper, are located in SOM.
- 23. A description of these studies is located in SOM.
- A. Zahavi, Mate selection-a selection for a handicap. J. Theor. Biol. 53, 205 (1975). doi:10.1016/0022-5193(75)90111-3 Medline
- A. Grafen, Biological signals as handicaps. J. Theor. Biol. 144, 517 (1990). doi:10.1016/S0022-5193(05)80088-8 Medline
- M. Andersson, Evolution of condition-dependent sex ornaments and mating preferences: Sexual selection based on viability differences. *Evolution* 40, 804 (1986). doi:10.2307/2408465
- P. D. Lorch, S. Proulx, L. Rowe, T. Day, Condition-dependent sexual selection can accelerate adaptation. *Evol. Ecol. Res.* 5, 867 (2003).
- J. L. Tomkins, J. Radwan, J. S. Kotiaho, T. Tregenza, Genic capture and resolving the lek paradox. *Trends Ecol. Evol.* **19**, 323 (2004). <u>doi:10.1016/j.tree.2004.03.029 Medline</u>
- 29. G. A. Parker, Assessment strategy and the evolution of fighting behaviour. J. Theor. Biol. 47, 223 (1974). doi:10.1016/0022-5193(74)90111-8 Medline
- M. Enquist, O. Leimar, Evolution of fighting behavior: Decision rules and assessment of relative strength. J. Theor. Biol. 102, 387 (1983). doi:10.1016/0022-5193(83)90376-4
- A. Pomiankowski, The handicap principle does work sometimes. Proc. R. Soc. Lond. B Biol. Sci. 231, 123 (1987). doi:10.1098/rspb.1987.0038
- R. Bonduriansky, T. Day, The evolution of static allometry in sexually selected traits. *Evolution* 57, 2450 (2003). <u>Medline</u>
- A. Kodric-Brown, R. M. Sibly, J. H. Brown, The allometry of ornaments and weapons. *Proc. Natl. Acad. Sci. U.S.A.* **103**, 8733 (2006). <u>doi:10.1073/pnas.0602994103</u> <u>Medline</u>
- J. S. Kotiaho, Costs of sexual traits: A mismatch between theoretical considerations and empirical evidence. *Biol. Rev. Camb. Philos. Soc.* 76, 365 (2001). doi:10.1017/S1464793101005711 Medline
- 35. J. F. Husak, J. G. Swallow, Compensatory traits and the evolution of male ornaments. *Behaviour* 148, 1 (2011). <u>doi:10.1163/000579510X541265</u>
- 36. In principle, selection on poor quality males to cheat could lead to evolutionary modifications to the underlying developmental mechanism that buffered expression of the exaggerated trait from the influence of male condition (i.e., that decreased sensitivity to insulin/IGF signals). In this event, the condition-sensitivity of trait expression and among-male variability in trait size would decrease (as in male genitalia of these beetles), reducing the reliability of the size of the trait as a signal of male quality. Interestingly, we are aware of no instances where exaggerated sexually selected signal traits presently display condition-*ins*ensitivity and/or reduced among-individual variation. This could be because once the traits become exaggerated, their costs reinforce signal honesty and select against cheating males. Or it could reflect the fact that once subsequent insensitivity to insulin/IGF evolves in an exaggerated trait, its reliability as a signal plummets, favoring receivers who ignore the trait and focus instead on other signals.
- C. E. Oufiero, T. Garland Jr., Evaluating performance costs of sexually selected traits. *Funct. Ecol.* 21, 676 (2007). doi:10.1111/j.1365-

2435.2007.01259.x

- A. R. Saltiel, C. R. Kahn, Insulin signalling and the regulation of glucose and lipid metabolism. *Nature* 414, 799 (2001). doi:10.1038/414799a Medline
- M. Tatar, A. Bartke, A. Antebi, The endocrine regulation of aging by insulinlike signals. *Science* 299, 1346 (2003). doi:10.1126/science.1081447 Medline
- 40. L. Fontana, L. Partridge, V. D. Longo, Extending healthy life span—from yeast to humans. *Science* 328, 321 (2010). <u>doi:10.1126/science.1172539</u> <u>Medline</u>
- B. A. Edgar, How flies get their size: Genetics meets physiology. Nat. Rev. Genet. 7, 907 (2006). doi:10.1038/nrg1989 Medline
- 42. J. Nakae, Y. Kido, D. Accili, Distinct and overlapping functions of insulin and IGF-I receptors. *Endocr. Rev.* 22, 818 (2001). <u>doi:10.1210/er.22.6.818</u> <u>Medline</u>
- 43. S. Oldham, E. Hafen, Insulin/IGF and target of rapamycin signaling: A TOR de force in growth control. *Trends Cell Biol.* 13, 79 (2003). doi:10.1016/S0962-8924(02)00042-9 Medline
- L. A. Johnston, P. Gallant, Control of growth and organ size in *Drosophila*. *Bioessays* 24, 54 (2002). <u>doi:10.1002/bies.10021</u> Medline
- A. A. Teleman, Molecular mechanisms of metabolic regulation by insulin in Drosophila. Biochem. J. 425, 13 (2010). doi:10.1042/BJ20091181 Medline
- 46. I. Claeys *et al.*, Insulin-related peptides and their conserved signal transduction pathway. *Peptides* 23, 807 (2002). <u>doi:10.1016/S0196-9781(01)00666-0 Medline</u>
- 47. Q. Wu, M. R. Brown, Signaling and function of insulin-like peptides in insects. Annu. Rev. Entomol. 51, 1 (2006). doi:10.1146/annurev.ento.51.110104.151011 Medline
- J. Baker, J.-P. Liu, E. J. Robertson, A. Efstratiadis, Role of insulin-like growth factors in embryonic and postnatal growth. *Cell* **75**, 73 (1993). <u>Medline</u>
- J. P. McMurtry, G. L. Francis, Z. Upton, Insulin-like growth factors in poultry. *Domest. Anim. Endocrinol.* 14, 199 (1997). <u>doi:10.1016/S0739-7240(97)00019-2 Medline</u>
- 50. T. Ventura *et al.*, Expression of an androgenic gland-specific insulin-like peptide during the course of prawn sexual and morphotypic differentiation. *ISRN Endocr.* Article ID 476283 (2011).
- D. Weinkove, S. J. Leevers, The genetic control of organ growth: Insights from *Drosophila. Curr. Opin. Genet. Dev.* 10, 75 (2000). <u>doi:10.1016/S0959-</u> 437X(99)00042-8 <u>Medline</u>
- O. Puig, R. Tjian, Transcriptional feedback control of insulin receptor by dFOXO/FOXO1. Genes Dev. 19, 2435 (2005). doi:10.1101/gad.1340505 Medline
- T. Ikeya, M. Galic, P. Belawat, K. Nairz, E. Hafen, Nutrient-dependent expression of insulin-like peptides from neuroendocrine cells in the CNS contributes to growth regulation in *Drosophila. Curr. Biol.* **12**, 1293 (2002). doi:10.1016/S0960-9822(02)01043-6 Medline
- W. Brogiolo *et al.*, An evolutionarily conserved function of the *Drosophila* insulin receptor and insulin-like peptides in growth control. *Curr. Biol.* 11, 213 (2001). <u>doi:10.1016/S0960-9822(01)00068-9</u> <u>Medline</u>
- P. J. Bryant, Growth factors controlling imaginal disc growth in *Drosophila*. Nov. Found. Symp. 237, 182 (2001).
- F. Lupu, J. D. Terwilliger, K. Lee, G. V. Segre, A. Efstratiadis, Roles of growth hormone and insulin-like growth factor 1 in mouse postnatal growth. *Dev. Biol.* 229, 141 (2001). doi:10.1006/dbio.2000.9975 Medline
- J. Karpac, H. Jasper, Insulin and JNK: Optimizing metabolic homeostasis and lifespan. *Trends Endocrinol. Metab.* 20, 100 (2009). doi:10.1016/j.tem.2008.11.004 Medline
- M. S. Dionne, L. N. Pham, M. Shirasu-Hiza, D. S. Schneider, *Akt* and *FOXO* dysregulation contribute to infection-induced wasting in *Drosophila*. *Curr. Biol.* 16, 1977 (2006). doi:10.1016/j.cub.2006.08.052 Medline
- 59. J. R. DiAngelo, M. L. Bland, S. Bambina, S. Cherry, M. J. Birnbaum, The immune response attenuates growth and nutrient storage in *Drosophila* by reducing insulin signaling. *Proc. Natl. Acad. Sci. U.S.A.* **106**, 20853 (2009). <u>doi:10.1073/pnas.0906749106 Medline</u>
- A. W. Shingleton, C. M. Estep, M. V. Driscoll, I. Dworkin, Many ways to be small: Different environmental regulators of size generate distinct scaling relationships in *Drosophila melanogaster*. *Proc. Biol. Sci.* 276, 2625 (2009). doi:10.1098/rspb.2008.1796 Medline
- B. T. Shea, R. E. Hammer, R. L. Brinster, Growth allometry of the organs in giant transgenic mice. *Endocrinology* **121**, 1924 (1987). <u>doi:10.1210/endo-121-6-1924 Medline</u>

- S. J. Leevers, D. Weinkove, L. K. MacDougall, E. Hafen, M. D. Waterfield, The *Drosophila* phosphoinositide 3-kinase Dp110 promotes cell growth. *EMBO J.* 15, 6584 (1996). <u>Medline</u>
- H. Huang *et al.*, PTEN affects cell size, cell proliferation and apoptosis during Drosophila eye development. Development **126**, 5365 (1999). <u>Medline</u>
- 64. D. Weinkove, T. P. Neufeld, T. Twardzik, M. D. Waterfield, S. J. Leevers, Regulation of imaginal disc cell size, cell number and organ size by *Drosophila* class I<sub>A</sub> phosphoinositide 3-kinase and its adaptor. *Curr. Biol.* 9, 1019 (1999). doi:10.1016/S0960-9822(99)80450-3 Medline
- 65. B. R. Beckman, M. Shimizu, B. A. Gadberry, K. A. Cooper, Response of the somatotropic axis of juvenile coho salmon to alterations in plane of nutrition with an analysis of the relationships among growth rate and circulating IGF-I and 41 kDa IGFBP. *Gen. Comp. Endocrinol.* **135**, 334 (2004). <u>doi:10.1016/j.ygcen.2003.10.013 Medline</u>
- 66. H. Charniaux-Cotton, C. Zerbib, J. J. Meusy, Monographie de la glande androgène des Crusacés supérieurs. *Crustaceana* 10, 113 (1966). <u>doi:10.1163/156854066X00658</u>
- 67. C. Nagamine, A. W. Knight, A. Maggenti, G. Paxman, Masculinization of female *Macrobrachium rosenbergii* (de Man) (Decapoda, Palaemonidae) by androgenic gland implantation. *Gen. Comp. Endocrinol.* **41**, 442 (1980). <u>doi:10.1016/0016-6480(80)90049-0 Medline</u>
- D. J. Emlen, Q. Szafran, L. S. Corley, I. Dworkin, Insulin signaling and limbpatterning: Candidate pathways for the origin and evolutionary diversification of beetle 'horns'. *Heredity* 97, 179 (2006). <u>doi:10.1038/sj.hdy.6800868</u> <u>Medline</u>
- 69. J. M. Suttie *et al.*, Insulin-like growth factor 1 (IGF-1) antler-stimulating hormone? *Endocrinology* **116**, 846 (1985). <u>doi:10.1210/endo-116-2-846</u> <u>Medline</u>
- J. M. Suttie, I. D. Corson, P. D. Gluckman, P. F. Fennessy, Insulin-like growth factor 1, growth and body composition in red deer stags. *Anim. Prod.* 53, 237 (1991). <u>doi:10.1017/S0003356100020171</u>
- 71. J. L. Elliott, J. M. Oldham, G. W. Asher, P. C. Molan, J. J. Bass, Effect of testosterone on binding of insulin-like growth factor-I (IGF-I) and IGF-II in growing antlers of fallow deer (*Dama dama*). *Growth Regul.* 6, 214 (1996). <u>Medline</u>
- 72. L. Gu *et al.*, Expression and localization of insulin-like growth factor-I in four parts of the red deer antler. *Growth Factors* 25, 264 (2007). <u>doi:10.1080/08977190701773187</u> Medline
- M. Sadighi, S. R. Haines, A. Skottner, A. J. Harris, J. M. Suttie, Effects of insulin-like growth factor-I (IGF-I) and IGF-II on the growth of antler cells in vitro. *J. Endocr.* 143, 461 (1994). doi:10.1677/joe.0.1430461 Medline
- 74. J. S. Price, B. O. Oyajobi, R. O. C. Oreffo, R. G. Russell, Cells cultured from the growing tip of red deer antler express alkaline phosphatase and proliferate in response to insulin-like growth factor-I. J. Endocrinol. 143, R9 (1994). <u>doi:10.1677/joe.0.143R009</u>
- D. J. Emlen, Environmental control of horn length dimorphism in the beetle Onthophagus acuminathus (Coleoptera: Scarabaeidae). Proc. Biol. Sci. 256, 131 (1994). doi:10.1098/rspb.1994.0060
- Y. Iguchi, Horn dimorphism in Allomyrina dichotoma septentrionalis (Coleoptera: Scarabaeidae) affected by larval nutrition. Ann. Entomol. Soc. Am. 91, 845 (1998).
- J. Hunt, L. W. Simmons, Maternal and paternal effects on offspring phenotype in the dung beetle *Onthophagus taurus*. *Evolution* 54, 936 (2000). <u>Medline</u>
- K. Karino, N. Seki, M. Chiba, Larval nutritional environment determines adult size in Japanese horned beetles *Allomyrina dichotoma. Ecol. Res.* 19, 663 (2004). doi:10.1111/j.1440-1703.2004.00681.x
- 79. K. Karino, H. Niiyama, M. Chiba, Horn length is the determining factor in the outcomes of escalated fights among male Japanese horned beetles, *Allomyrina dichotoma* L. (Coleoptera: Scarabaeidae). J. Insect Behav. 18, 805 (2005). <u>doi:10.1007/s10905-005-8741-5</u>
- Y. Hongo, Evolution of male dimorphic allometry in a population of the Japanese horned beetle *Trypoxylus dichotomus septentrionalis*. *Behav. Ecol. Sociobiol.* 62, 245 (2007). doi:10.1007/s00265-007-0459-2
- 81. J. Lai, K. Shin-Ping, For the Love of Rhinoceros and Stag Beetles (2008).
- M. D. Abramoff, P. J. Magalhaes, S. J. Ram, Image Processing with ImageJ. Biophotonics Int. 11, 36 (2004).
- E. Roovers *et al.*, Characterization of a putative molluscan insulin-related peptide receptor. *Gene* 162, 181 (1995). <u>doi:10.1016/0378-1119(95)00323-X</u>

Medline

- Y. Tomoyasu, R. E. Denell, Larval RNAi in *Tribolium* (Coleoptera) for analyzing adult development. *Dev. Genes Evol.* 214, 575 (2004). doi:10.1007/s00427-004-0434-0 Medline
- J. Sambrook, W. R. Russel, *Molecular Cloning, A Laboratory Manual*. (Cold Spring Harbor Laboratory Press, Cold Spring Harbour 2001).
- J. Hellemans, G. Mortier, A. De Paepe, F. Speleman, J. Vandesompele, qBase relative quantification framework and software for management and automated analysis of real-time quantitative PCR data. *Genome Biol.* 8, R19 (2007). doi:10.1186/gb-2007-8-2-r19 Medline
- W. F. Marzluff, E. J. Wagner, R. J. Duronio, Metabolism and regulation of canonical histone mRNAs: Life without a poly(A) tail. *Nat. Rev. Genet.* 9, 843 (2008). doi:10.1038/nrg2438 Medline
- D. I. Warton, I. J. Wright, D. S. Falster, M. Westoby, Bivariate line-fitting methods for allometry. *Biol. Rev. Camb. Philos. Soc.* 81, 259 (2006). doi:10.1017/S1464793106007007 Medline
- R. J. Nelson, Introduction to Behavioral Endocrinology. (Sinauer, Sunderland, Mass 2011).
- C. Mirth, J. W. Truman, L. M. Riddiford, The role of the prothoracic gland in determining critical weight for metamorphosis in *Drosophila melanogaster*. *Curr. Biol.* 15, 1796 (2005). <u>doi:10.1016/j.cub.2005.09.017 Medline</u>
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### Supplementary Materials

www.sciencemag.org/cgi/content/full/science.1224286/DC1 Materials and Methods Supplementary Text Figs. S1 to S3 Tables S1 and S2 References (*38–90*)

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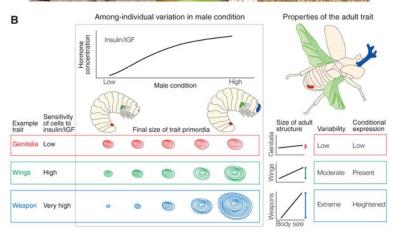
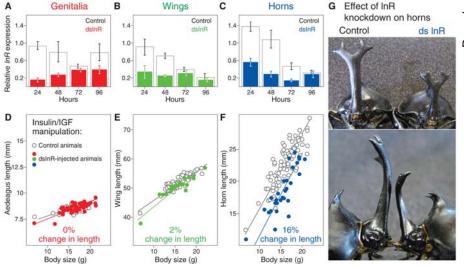


Fig. 1. A). Exaggerated growth of weapons and ornaments of sexual selection as exemplified by rhinoceros beetle horns (Trypoxylus dichotomus). B) Proposed mechanism for the evolution of trait exaggeration through increased cellular sensitivity to insulin/IGF signaling (shown for the disc-like appendage primordia of insects). Individual nutritional state and physiological condition are reflected in circulating levels of insulin-like peptides and IGFs, which modulate the rate of growth of each of the trait primordia. Traits whose cells are sensitive (17) to these signals (e.g., wings [green]) exhibit greater nutrition-dependent phenotypic plasticity and among-individual variability than other traits whose cells are less sensitive to these signals (e.g., genitalia [red]). An increase in the sensitivity of cells within a particular trait (e.g., horns [blue], see text) would lead to disproportionately rapid growth of that trait in the largest, best-condition individuals (i.e., exaggerated trait size) and smaller trait sizes in low-condition individuals.

Fig. 2. Effect of insulin receptor (InR) knockdown on growth of adult structures in rhinoceros beetles. A-C) Relative transcript abundances for the insulin receptor (InR) gene in genitalia (A), wings (B), and horns (C), measured 24, 48, 72, and 96 hours after the onset of the prepupal period in control (open bars) and dsInR-injected (solid bars) animals. Injection with dsRNA significantly reduced transcript abundances for 48 hours following injection in all three tissues. D-F) Effects of dsInR knockdown on trait growth. Genitalia were insensitive (D); responded wings significantly but moderately to interrupted insulin/IGF signaling (E) (average reduction in wing length = 2%); and horns responded dramatically (F), with an average reduction in horn length of 16%. G. Head and thorax shown in two orientations (top and bottom) for same-sized control (left) and dsInR-injected (right) males.



А

Category of models:	Model conclusions:	Model predictions:	Contribution of the insulin/IGF mechanism:
Corroborated by an insulin/IGF	mechanism for trait exaggeration:		
Index signal models Maynard Smith & Harper 2003 ( <i>3</i> )	Sexually selected traits can evolve to exaggerated sizes if bigger versions of the trait are more effective signals than smaller versions, and if trait size is a physiologically or physically unfakable 'index' of the quality of the signaler. Trait size is a reliable signal because it is mechanistically impossible for low quality signalers to cheat.	Should be mechanistically impossible for low quality males to produce large signal traits. Why the size of a signal trait is unfakable is often unclear/unspecified.	Provides an explicit mechanism for unfakable signal expression in exaggerated morphological structures.
Good genes/indicator models Andersson 1986 (26) Iwasa & Pomiankowski 1999 (13) Lorch et al. 2003 (27)	Females benefit if they choose mates based on conditionally expressed ornaments because variation in the expression of these traits reliably indicates the overall quality of a male. Low quality males produce smaller signal traits because they are in poor condition.	Exaggerated traits should exhibit 'heightened' conditional expression.	Provides an explicit mechanism for heightened conditional expression of exaggerated morphological structures.
Genic capture models Rowe & Houle 1996 (1/2) Lorch et al. 2003 (27) Tomkins et al. 2004 (28)	Evolution of ornaments persists (genetic variation is not depleted) because in their expression these traits "capture" genetic variation for overall body condition, including health, resistance to parasites, competitive ability, nutriton, etc.	Genetic variation among males affecting their body condition, resistance to parasites, competitive ability, etc., should translate into differences in ornament size.	Provides an explicit mechanism for genic capture, since all of these aspects of body condition are channeled into a common endocrine signal regulating trait growth.
Assessment/arms race models Parker 1974 ( <i>29</i> ) Enquist & Leimar 1983 ( <i>30</i> )	Male weapons can evolve to exaggerated sizes if weapon size reliably signals the fighting ability of a male.	Males should use relative weapon size as a basis for assessment; fights should be most likely to escalate if rival males are similarly armed. Not clear from the models why weapon size should remain reliable.	Suggests that weapons will become increasingly reliable signals of fighting ability as they increase in size, facilitating arms races.
Modified by an insulin/IGF med	chanism for trait exaggeration:		
Handicap models Zahavi 1975 (24) Pomiankowski 1987 (31) Grafen 1990 (25) Johnstone 1995 (11) Iwasa & Pomiankowski 1999 (13)	Females benefit if they choose mates based on costly ornaments. A given increase in trait size costs low quality males more (or benefits them less) than it does high quality males, resulting in ornament sizes that reliably signal male quality.	Costs should be present, and they should be relatively highest for low quality males.	Suggests that costs may not be necessary for maintaining signal reliability <sup>2</sup> , and the handicap principle may only be relevant when exaggeration is extreme.
Allometry evolution models Bonduriansky & Day 2003 (32) Kodric-Brown et al. 2006 (33)	Exaggerated ornaments/ weapons will have steep allometry slopes when small males pay higher costs (or derive fewer benefis) than large males for the same increase in trait size.	Costs of ornaments/weapons should trade-off with allocation to overall growth or body mainte- nance, and these costs should be relatively highest for small males.	Suggests that costs may not be necessary for steep allometry slopes (they should arise as a byproduct of the mechanism of exaggeration)*. This should expand the conditions for which steep allometry slopes are expected.

**Fig. 3.** Sexual selection models whose relevance is affected by the proximate mechanism responsible for trait exaggeration.